

Goal

This study aims to improve localized sampling of small areas of FFPE and fresh-frozen tissues and uncover proteomic differences between the two conservation techniques for improved analytical sensitivity.

Introduction

- Microsampling for proteomics is reproducible with laser ablation
- Micro-volume sample processing for small area analysis
- FFPE capabilities improve scope of method
- FFPE tissue analysis difficult due to protein crosslinking
- New workflow needed for FFPE tissue microsampling

Experimental



≻IR laser:

- Optical parametric oscillator (OPO)
- 2.94 µm
- 300 kJ/m² fluence

> Microwell collection:

- DMSO-containing PTFE-coated slide
- 2 mm diameter well
- \geq Rat brain tissue section:
- 10 µm thick

Infrared Laser Ablation Microsampling for Small Volume Proteomics of Formalin Fixed Paraffin Embedded Tissue B. Chisom Egbejiogu; Fabrizio Donnarumma; Kermit K. Murray Louisiana State University, Department of Chemistry, Baton Rouge, LA





PTFE printed glass microscope

efficiency with minimal solvent evaporation.

Each ablated tissue region was collected and digested in situ prior to LC-MS/MS analysis.

Protein/Peptide Identification

➢ Region: Midbrain

 \succ Reproducible protein identification with an average CV <12% for # of peptides and proteins



Gene ontology analysis (Panther GO) of all proteins identified for each

- Efficient ablation and capture from both FFPE and FF tissue
- >400 proteins and >900 peptides identified from mm² area
- Comparable cellular components and biological processes FFPE vs. FF

Conclusions

- Simple micro-volume workup adapted for FFPE microsampling
- Efficient proteomics down to 0.4 mm² area containing <5 ng protein

Future work

- Sample clinically relevant tissue (FFPE cancer biopsies)
- Sample smaller areas of FF and FFPE tissue
- Laser ablation sampling of stained tissue \bullet

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AVERAGE TOTAL









Percentage

Biological Processes